9/7639 T/AU99/00705 AU99/705

REC'D 1 3 OCT 1999 PCT WIPO

Patent Office Canberra

I, LEANNE MYNOT, TEAM LEADER EXAMINATION SUPPORT AND SALES hereby certify that annexed is a true copy of the Provisional specification in connection with Application No. PP 5572 for a patent by THE UNIVERSITY OF QUEENSLAND filed on 31 August 1998.



WITNESS my hand this Sixth day of October 1999

LEANNE MYNOTT

TEAM LEADER EXAMINATION SUPPORT AND SALES

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PROVISIONAL SPECIFICATION

for the invention entitled:

"A novel plant promoter and uses therefor"

The invention is described in the following statement:

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Plants are subject to a variety of environmental and mechanical stimuli including stress. Although mechanical stress has been postulated to involve ethylene-mediated meristem morphogenesis (Selker *et al*, 1992), little is known about how mechanical stress induces ethylene production or the signal transduction process involved.

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In work leading to the present invention, the inventors sought to identify and isolate promoters involved in mechanical stress-induced expression of genetic traits in *Vigna radiata* (mung bean). Mung bean plants are a useful model for physical and chemical induction of phenotypic expression of genetic traits due to their morphology, rapid growth rate and the ability to obtain a large number of uniform plants and, therefore, sufficient amounts of tissues to conduct analyses.

In accordance with the present invention, the inventors have isolated a promoter capable of induction following physical stimulus in cells in which the promoter is indigenous, i.e. cells of mung bean plants. The promoter is also capable of being induced by a range of chemical and other environmental stimuli. However, in cells in which the promoter is non-indigenous, the promoter is constitutively expressed. The promoter of the present invention is useful in the genetic manipulation of plants.

20 SUMMARY OF THE INVENTION

Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated element or integer or group of elements or integers but not the exclusion of any other element or integer or group of elements or integers.

One aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein, in its native form, the promoter is inducible in response to physical stimulation.

Another aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof, wherein said promoter, in its native form, directs expression of a gene associated with ethylene production and is inducible by physical stimulation.

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Yet another aspect of the present invention relates to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding ACC synthase and is inducible by physical stimulation.

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Still another aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding ACC synthase or another gene associated with ethylene biosynthesis and is inducible by physical stimulation.

Still yet another aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding an ACC synthase having an amino acid sequence substantially as set forth in SEQ ID NO:2 or an amino acid sequence having at least 60% similarity to SEQ ID

NO:2.

A further aspect of the present invention relates to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding ACC synthase and wherein said gene comprises a nucleotide sequence substantially as set forth in SEQ ID NO:1 or a sequence having at least 50% similarity thereto and/or a nucleotide sequence capable of hybridizing to SEQ ID NO:1 under low stringency conditions at 42°C.

Still another aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, comprises a nucleotide sequence substantially as set forth in SEQ ID NO:3 or a nucleotide sequence having at least 25% similarity thereto or a nucleotide sequence capable of hybridizing to SEQ ID NO:3 under low stringency conditions at 42°C.

Another aspect of the present invention provides a modular promoter, said modular promoter comprising at least one portion which is derived from a promoter which, in its native form, directs expression of a gene associated with ethylene biosynthesis and is inducible by physical stimulation.

Yet another aspect of the present invention is directed to a modular promoter, said modular promoter comprising at least one portion which is derived from a promoter which, in its native form, directs synthesis of an ACC synthase having an amino acid sequence substantially as set forth in SEQ ID NO:2 or an amino acid sequence having at least 60% similarity thereto.

Still yet another aspect of the present invention is directed to a modular promoter, said modular promoter comprising at least one portion which is derived from a promoter which, it is native form, directs synthesis of an ACC synthase encoded by a gene comprising a nucleotide sequence substantially as set forth in SEQ ID NO:1 or a nucleotide sequence having at least 50% similarity thereto or a nucleotide sequence capable of hybridizing to SEQ ID NO:1 under low stringency conditions at 42°C.

25 In still yet another aspect of the present invention, there is provided a modular promoter comprising a portion which is derived from a promoter which comprises, in its native form, a nucleotide sequence substantially as set forth in SEQ ID NO:3 or a nucleotide sequence having at least 25% similarity thereto or a nucleotide sequence capable of hybridizing to SEQ ID NO:3 under low stringency conditions at 42°C.

or modular promoter each as herein defined or a derivative or homologue thereof, means to facilitate insertion of a nucleotide sequence downstream of and operably linked to said promoter and optionally a gene encoding a selectable marker.

5 A further aspect of the present invention provides a genetic construct comprising a promoter or modular promoter each as herein defined or a derivative or homologue thereof, a nucleotide sequence operably linked to said promoter and optionally a gene encoding a selectable marker.

Another aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof which is capable of constitutive expression in cells in which the promoter is non-indigenous.

Yet another aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof, wherein said promoter, in its native form, directs expression in response to physical stimulation of a gene associated with ethylene production and which promoter in a non-native host cell is constitutively expressed.

20 Still yet another aspect of the present invention relates to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression in response to physical stimulation of a gene encoding ACC synthase and which promoter in a non-native host cell is constitutively expressed.

25

A further aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding ACC synthase or another gene associated with ethylene biosynthesis and in a cell in which the promoter is indigenous, the promoter is inducible by physical stimulation whereas in a cell in which the promoter is non-indigenous, the promoter is constitutively

expressed.

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BRIEF DESCRIPTION OF THE FIGURES

- 5 Figure 1 is a representation of the oligonucleotide primers used in Long Distance Inverse PCR.
 - Figure 2 is a diagrammatic representation showing generation of *SpeI* and *Xba-I* fragments of *AIM-I* promoter.
- 10 Figure 3 is a diagrammatic representation of the AIM-1 promoter sequencing strategy.
 - Figure 4 is a representation of the nucleotide sequence of the AIM-1 (2470 bp) promoter.
 - Figure 5 is a diagrammatic representation of the construction of a full length AIM-1 promoter.
 - Figure 6A(i) 6A(xii) are diagrammatic representations of plasmids pPZP2.5GuNt, pPZP2.5LuNt, pPZP1.4GuNt, pPZP1.4LuNt, pPZP35SGuNt, pPZP35SLuNt, pPZP017GuNt, pPZP023GuNt, pPZP07GuNt, pPZP045GuNt, pPZP088GuNt and pPZP1.1GuNt, respectively.
- 20 Figure 6B is a diagrammatic representation of the backbone vector pPZP111.
 - Figure 6C is a diagrammatic representation of the vector pGuNt.
- Figure 7 are photographic representations showing (A) and (B) transgenic tobacco lines containing AIM-1:GUS gene assayed to visualise GUS activity; and (C) non-stained controls.
- Figures 8(A) and (B) are graphical representations showing GUS activity in young tobacco plants, transformed with AIM-1:GUS and CaMV35S:GUS constructs. 2.5G#3-4 and 2.5G#7-3 are two independent transgenic lines containing full length AIM-1 promoter fused to the GUS gene; 35SG#5-2 is a transgenic line containing CaMV35S promoter fused to the GUS gene. (A) is GUS activity per/mg protein. (B) is GUS activity per/g of fresh weight. MU is equal to 4-

methyl-umbrelliferone.

Figure 9 is a photographic representation of Southern analysis of three T2 homozygous independent tobacco transgenic lines (3-4, 7-3 and 10-3) containing the AIM-1 promoter fused to the GUS gene, and one T2 homozygous tobacco transgenic line (5-2) containing the CaMV 35S promoter fused to the GUS gene. Genomic DNA was digested with EcoRI (E) or BamHI (B) restriction enzymes. A ³²P-labelled DNA fragment containing the full GUS gene and Nos terminator was used as a probe. Lane 1 contained size markers. Lanes 2 and 3: line 3-4; lanes 4 and 5: line 7-3; lanes 6 and 7: line 10-3; lane 8: line 5-2.

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DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is predicated in part on the identification of a promoter directing expression of a gene. The gene encodes 1-aminocyclopropane-1-carboxylic acid synthase ("ACC synthase") and is inducible, in its native form, by physical stimuli (Botella *et al*, 1992; Botella *et al*, 1995). Referenced herein to "native form" with respect to a promoter means the promoter in cells in which the promoter is normally resident, i.e. indigenous. In the present case, cells for mung bean plants are cells in which the promoter is indigenous. Transfer of the promoter by genetic means to other plant cells is an example of cells carrying a non-indigenous promoter.

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Accordingly, the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein, in its native form, the promoter is inducible in response to physical stimulation.

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More particularly, the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof, wherein said promoter, in its native form, directs expression of a gene associated with ethylene production and inducible by physical stimulation.

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Even more particularly, the present invention relates to an isolated nucleic acid molecule

comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding ACC synthase and inducible by physical stimulation.

5 In a related embodiment, the present invention relates to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding ACC synthase or another gene associated with ethylene biosynthesis and is inducible by physical stimulation.

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Although the present invention is exemplified by the identification and isolation of the promoter directing synthesis of ACC synthase from Vigna radiata (mung bean), the present invention extends to any promoter which, in its native form, is inducible in response to physical stimulation and which comprises a nucleotide sequence having at least about 50% similarity to the nucleotide sequence set forth in SEQ ID NO:1 and/or nucleotide sequence capable of hybridizing to the nucleotide sequence of SEQ ID NO:1 under low stringency conditions at 42°C.

Examples of promoters contemplated by the present invention include but are not limited to promoters directing expression of genes associated with ethylene biosynthesis such as the gene 20 encoding ACC synthase.

The gene encoding ACC synthase from mung bean is referred to as "AIM-1". ACC synthase from mung bean comprises the amino acid sequence substantially as set forth in SEQ ID NO:2.

25 Accordingly, another aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding an ACC synthase having an amino acid sequence substantially as set forth in SEQ ID NO:2 or an amino acid sequence having at least 60% 30 similarity to SEQ ID NO:2.

The percentage similarity at the amino acid or nucleotide sequence level is generally to a portion comprising at least about 20 contiguous amino acids or at least about 60 contiguous nucleotide bases. Preferably, however, the comparison is made to the entire amino acid sequence or entire nucleotide sequence. Alternative percentage similarities include at least about 70%, at least about 80%, at least about 90% and at least about 95% or above or discrete percentages there between.

Genes encoding ACC synthase enzymes not having 100% similarity to SEQ ID NO:2 include derivatives and homologous of the mung bean enzyme. A derivative includes parts, fragments, mutants and fusions of the mung bean ACC synthase defined in SEQ ID NO:2 including ACC synthase enzymes having one or more amino acid substitutions, additions and/or deletions to the amino acid sequence of SEQ ID NO:2. Homologues include enzymes from closely or distantly related plants including fungi.

15 A particularly preferred promoter of the present invention directs expression of AIM-1. The nucleotide sequence of AIM-1 is set forth in SEQ ID NO:1.

According to this embodiment, there is provided an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter in its native form directs expression of a gene encoding ACC synthase and wherein said gene comprises a nucleotide sequence substantially as set forth in SEQ ID NO:1 or a sequence having at least 50% similarity thereto and/or a nucleotide sequence capable of hybridizing to SEQ ID NO:1 under low stringency conditions at 42°C.

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Reference herein to a low stringency at 42°C includes and encompasses from at least about 1% v/v to at least about 15% v/v formamide and from at least about 1M to at least about 2M salt for hybridisation, and at least about 1M to at least about 2M salt for washing conditions. Alternative stringency conditions may be applied where necessary, such as medium stringency, which includes and encompasses from at least about 16% v/v to at least about 30% v/v formamide and from at least about 0.5M to at least about 0.9M salt for hybridisation, and at least about 0.5M

to at least about 0.9M salt for washing conditions, or high stringency, which includes and encompasses from at least about 31% v/v to at least about 50% v/v formamide and from at least about 0.01M to at least about 0.15M salt for hybridisation, and at least about 0.01M to at least about 0.15M salt for washing conditions.

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Alternative percentage similarities include those set forth above.

Nucleotide sequences not having 100% similarity to SEQ ID NO:1 include derivatives and homologues of mung bean AIM-1. A derivative includes, parts, fragments, mutants and fusions of the mung bean AIM-1 defined in SEQ ID NO:1 including AIM-1 genes having one or more nucleotide substitutions, additions and/or deletions to the nucleotide sequence of SEQ ID NO:1. Homologues include genes from closely or distantly related plants including fungi.

The term "similarity" as used herein includes exact identity between compared sequences at the nucleotide or amino acid level. Where there is non-identity at the nucleotide level, "similarity" includes differences between sequences which result in different amino acids that are nevertheless related to each other at the structural, functional, biochemical and/or conformational levels. Where there is non-identity at the amino acid level, "similarity" includes amino acids that are nevertheless related to each other at the structural, functional, biochemical and/or conformational levels. The terms "homology" and "identity" may be used to substitute for "similarity".

Most preferably, the promoter of the present invention comprises a nucleotide sequence substantially as set forth in SEQ ID NO:3 or a functional derivative or homologue thereof.

25 Accordingly, another aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter in its native form comprises a nucleotide sequence substantially as set forth in SEQ ID NO:3 or a nucleotide sequence having at least 25% similarity thereto or a nucleotide sequence capable of hybridizing 30 to SEQ ID NO:3 under low stringency conditions at 42°C.

The promoter of the present invention is useful in the development of genetic constructs to express heterologous nucleotide sequences placed downstream of, and operably linked to, the promoter.

5 The term "promoter" is used herein in its most general sense and refers to any nucleotide sequence which binds RNA polymerase and directs same to a transcriptional start site whereupon a gene or other nucleotide sequence downstream of said promoter is transcribed. A nucleotide sequence "downstream" of the promoter is also said to be "relative" to the promoter or "operably" linked to the promoter.

10

The term "genetic construct" is used in its broadest sense to include an isolated nucleic acid molecule comprising a sequence of nucleotides.

The genetic construct is conveniently engineered so as to include means to facilitate insertion of a nucleotide sequence in a region 3' of the promoter to place the nucleotide sequence downstream of and operably linked to the promoter for their transcription. Such a "means" includes but is not limited to a restriction endonuclease-mediated insertion, homologous recombination, transposon insertion, PCR mediated insertion and random insertion. Preferably, the means is a restriction endonuclease site. Generally, the inserted restriction site is unique to the genetic construct or may be represented, for example, twice but separated by a nucleic acid sequence which is deleted upon restriction digestion of the genetic construct. The required nucleotide sequence to be transcribed is then inserted into the deleted region.

The genetic construct of the present invention may comprise solely the promoter and optionally a nucleotide sequence downstream thereof or, alternatively, may comprise additional nucleotide sequences constituting promoter regulatory region(s), transcribed sequence regulatory regions, a marker (eg. antibiotic resistance, chemical compound resistance or enzyme), autonomous replication region and/or genome integration sequence. The promoter may be the naturally occurring promoter or may be an active fragment or part thereof or a derivative, analogue or

30 homologue of the promoter.

Accordingly, another aspect of the present invention contemplates a genetic construct comprising a promoter or modular promoter each as herein defined or a derivative or homologue thereof, means to facilitate insertion of a nucleotide sequence operably linked to said promoter and optionally a gene encoding a selectable marker.

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More particularly, this aspect provides a genetic construct comprising a promoter or modular promoter as herein defined or a derivative or homologue thereof, one or more unique restriction sites down stream of said promoter to enable the insertion of a heterologous nucleotide sequence operably linked to said promoter and a gene encoding a selectable marker.

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In a related embodiment, the present invention provides a genetic construct comprising a promoter or modular promoter each as herein defined or a derivative or homologue thereof, a nucleotide sequence operably linked to said promoter and optionally a gene encoding a selectable marker.

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The promoter of the present invention, in its native form (i.e. in cells in which it is indigenous), is inducible by physical stimulus which includes mechanical stress, movement, vibration, air pressure, water stress and the like. Other non-mechanical stimuli also induce the instant promoter including auxins, abscisic acid, salt concentration amongst others. Non-mechanical stimuli include environmental stimuli such as but not limited to chemical induction of the promoter. The promoter may also be developmentally regulated and/or may be tissue or organ specific.

The identification of a promoter capable of induction by physical or mechanical stimuli provides
25 a particularly useful basis for developing a range of genetically altered plants. For example, air
movement may be used to activate expression of a nucleotide sequence operably linked to the
subject promoter. This may be useful during the commercial cultivation of large numbers of
plants. Generating air movement such as generated by fanning, or a change in air pressure over
and/or around the plants can be used to activate expression of the promoter. Alternatively, or
30 in addition, water droplets generated mechanically or by controlling humidity may be used to

stimulate promoter activity. Heterologous nucleotide sequences operably linked to the promoter

are then expressed. Such heterologous sequences may encode, for example, resistance to insect or other pathogens, salt tolerance, enzymes which manipulate the flow of metabolites down particular biochemical pathways, enzymes which alter the nutritional content of certain types of plant tissues including seeds and other reproductive parts and antisense, co-suppression, ribozyme or deoxyribozyme molecules to down regulate expression of an endogenous gene. Examples of the latter would be to render a plant male or female sterile, to alter biochemical pathways or to otherwise alter the characteristics of the target plant, such as to inhibit ethylene biosynthesis or to delay synthesence.

10 Accordingly, another aspect of the present invention contemplates a method of altering a characteristic of a plant said method comprising introducing a genetic construct into a cell or group of cells of a plant, said genetic construct comprising a promoter as herein defined and a nucleotide sequence operably linked to said promoter and wherein said nucleotide sequence facilitates the altering of said plant characteristic, regenerating a plant or plantlet from said cell or group of cells carrying said genetic construct and growing or subjecting said plant or plantlet to conditions sufficient to induce the promoter in said genetic construct.

The genetically altered plant may be subjected to physical stimulus such as mechanical stress in order to induce the promoter. Alternative forms of stimulus, however, are also contemplated by the subject invention such as water droplets, air movement, air pressure and chemical stimuli such as auxins. The promoter may also be constitutively expressed.

An altered characteristic may be readily determined by comparing a transgenic plant with a non-transgenic plant of the same species. The comparison may be at the biochemical, physiological or visual level. Altered characteristics include but are not limited to resistance to plant viruses, bacteria, fungi, nematodes and other pathogens, improved nutritional value (eg. using sunflower high sulphur gene), an expression of an "antibody" (often referred to as a "plantabody"), altered biochemical pathways, altered fertility, altered flower colour amongst many other characteristics.

30 The promoter of the present invention is inducible by a range of stimuli including physical, environmental, chemical and genetic. The promoter comprises, therefore, different regulatory

areas for different stimuli. The present invention contemplates the manipulation of the subject promoter such that it is inducible by a particular stimulus or stimuli.

Accordingly, another aspect of the present invention provides a modular promoter, said modular promoter comprising at least one portion which is derived from a promoter which, in its native form, directs expression of a gene associated with ethylene biosynthesis and is inducible by physical stimulation.

More particularly, the present invention is directed to a modular promoter, said modular promoter comprising at least one portion which is derived from a promoter which, in its native form, directs synthesis of an ACC synthase having an amino acid sequence substantially as set forth in SEQ ID NO:2 or an amino acid sequence having at least 60% similarity thereto.

Even more particularly, the present invention is directed to a modular promoter, said modular promoter comprising at least one portion which is derived from a promoter which, it is native form, directs synthesis of an ACC synthase encoded by a gene comprising a nucleotide sequence substantially as set forth in SEQ ID NO:1 or a nucleotide sequence having at least 50% similarity thereto or a nucleotide sequence capable of hybridizing to SEQ ID NO:1 under low stringency conditions at 42°C.

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Still more particularly, the present invention provides a modular promoter comprising a portion which is derived from a promoter which comprises, in its native form, a nucleotide sequence substantially as set forth in SEQ ID NO:3 or a nucleotide sequence having at least 25% similarity thereto or a nucleotide sequence capable of hybridizing to SEQ ID NO:3 under low stringency conditions at 42°C.

A "modular" promoter is considered as an example of a "derivative". Another derivative contemplated by the present invention includes the deletion of negatively acting *cis* element(s). This aspect of the present invention is predicated on the observation of high expression of the promoter in the presence of the protein synthesis inhibitor, cycloheximide, which inhibits production of a highly unstable, short-lived negative regulator (transcription factor) of the

subject promoter. Accordingly, by deleting the negative *cis* element(s), higher inducible or even constitutive expression of the promoter may be obtained.

Another aspect of the present invention contemplates an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof which is capable of constitutive expression in cells in which the promoter is non-indigenous.

This aspect of the present invention is predicated on the surprising observation that the promoter of the present invention, when placed in plant cells in which it is not indigenous, i.e. non-mung bean cells, is constitutively expressed. Although no intending to limit the present invention to any one theory or mode of action, it is proposed that in cells in which the promoter is indigenous, a negative regulatory molecule prevents constitutive expression of the promoter. This negative regulatory molecule would not normally be present in other plant cells and, hence, the promoter is constitutively expressed.

Accordingly, another aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof, wherein said promoter, in its native form, directs expression in response to physical stimulation of a gene associated with ethylene production and in which in a non-native host cell is constitutively expressed.

More particularly, a further aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides or a complementary sequence of nucleotides defining a promoter or a derivative or homologue thereof wherein said promoter, in its native form, directs expression of a gene encoding ACC synthase or another gene associated with ethylene biosynthesis and in a cell in which the promoter is indigenous, the promoter is inducible by physical stimulation whereas in a cell in which the promoter is non-indigenous, the promoter is constitutively expressed.

The present invention further contemplates a transgenic plant carrying the promoter of the present invention or parts, limbs, flowers, petals, reproductive portions or seeds thereof or progeny or clone thereof.

5 The present invention is further described by the following non-limiting Examples.

EXAMPLE 1

Detection of mechanical strain-induced gene

10 A gene encoding 1-aminocyclopropane-1-carboxylic acid synthase ("ACC Synthase"), induced *inter alia* by mechanical strain, auxin and salt stress was isolated according to Botella *et al* (1992;1995). The cDNA sequence and corresponding amino acid sequence is shown in SEQ ID NO:1. The amino acid sequence alone is shown in SEQ ID NO:2.

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EXAMPLE 2

Cloning of the ACC Synthase gene (AIM-I) promoter

(a) Recirculation of DNA

Ten micrograms of genomic DNA isolated by CsCl purification was digested with 2.5U/µg of 20 HindIII in the presence of 0.1M spermidine, extracted with 1 volume phenol:chloroform:isamyl alcohol (25:24:1) and precipitated by addition of 0.1 vol NaOAc and 2 volumes EtOH. DNA was then re-ligated with 9 Weiss units of T4 DNA ligase and purified using Bresatec's Bresa Clean Kit. The effectiveness of recircularisation was analyzed by gel electrophoresis.

25

(b) Long Distance Inverse Polymerase Chain Reaction (LDIPCR) procedure

A reaction mixture of 2mM MgSO₄ pH 9.1, containing 60mM Tris-SO4, 18mM (NH₄) 2SO₄, 0.2mM of each dNTP, 0.2μM of NSE oligonucleotide primers (see Figure 1), sterile water and 300ng of recircularised genomic DNA was prepared in a total volume of 40μl. The reaction mixture was vortexed and briefly spun prior to incubation at 94°C to prevent non-

specific primer interactions. Before initialising the thermal cycle, 10µl of sterile water

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containing 1μ L of Life Technologies' eLONGase enzyme mix (Taql/Vent polymerases) was added to the reaction and mixed by pipetting. An equal volume of mineral oil was layered over the mix to prevent evaporation. The optimised PCR parameters are shown in Table 1.

TABLE 1

PCR profile times and temperatures used during amplification and reamplification protocols.

	Optimised Temperatures and Times											
		Initial Step	Denaturation	Anneal and Extension 480 sec.								
0	Amplification	60 sec.	30 sec.									
	Ampinication	94°C	94°C	94°C 68°C								
				45 cycles	·							
		Initial Step	Denaturation	Anneal	Extension							
	Reamplification	60 sec.	30 sec.	30 sec.	480 sec.							
	Reamphileation	94°C	94°C 62°C 68									
				35 cycles								

After the final step of thermal cycling, 1 volume of chloroform-isoamyl alcohol (24:1) was added to remove the oil layer and the samples were stored at 4°C.

20 Cloning Strategy

The circularised genomic DNA was first amplified with oligonucleotide primers NSE-1 and NSE-2 (refer to Figure 1). The products of this first amplification were further reamplified using either NSE-3/NSE-4 or NSE-5/NSE-6 (Figure 1). Electrophoretic analysis of the amplification products, generated with both combinations of primers, revealed a DNA

25 fragment of approximately 4kb.

EXAMPLE 3

Analysis of 4kb fragment

5 The 4kb product obtained with NSE3/NSE-4 (Example 2) was excised from the gel and purified with glassmilk (Bresatec's Bresa Clean). As attempts at cloning the 4kb product were initially unsuccessful, alternative strategies were devised. The purified 4kb product was digested with *Xba*l and two fragments of 1.3 kb and 0.9 kb were sub-cloned into the vector pGEM11 (Promega corporation, USA), which had been previously digested with *Xba*l giving the plasmids pGX1.3 and pGX0.9, respectively. The 4kb fragment was also digested with *Spe*I and blunt-ended before cloning the digestion products into pGEM11 (previously digested with *Xho*I and blunt-ended). As a result, two *Spe*I fragments of 1.1kb and 1.4kb were sub-cloned and the plasmids named pGS1.1 and pGS1.4, respectively. The 1.4kb fragment did not show any *Spe*I recognition sequences in one of its ends, indicating that some exonuclease activity had taken place during the blunt-ending process. This is summarised in Figure 2.

EXAMPLE 4

Reconstruction and sequencing of the 2.5kb AIM-I promoter region

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The sequencing strategy for the AIM-1 promoter is shown in Figure 3. Sequencing was performed using the dideoxy chain termination method (Sanger et al, 1977) using a Applied Biosystems kit (Applied Biosystems, USA). Analysis of the sequences revealed that the four clones partially overlapped. The 1.3 Xbal and 1.1kb SpeI fragments contained the 5'

- 25 untranslated region of the AIM-1 cDNA, confirming that this region is upstream of the AIM-1 gene. As a result, a partial restriction map for a 2.5kb region of the 4kb DNA fragment was generated. The nucleotide sequence of the AIM-1 promoter is shown in Figure 4 and in SEQ ID NO:3.
- 30 With this information in hand, the promoter region was reconstructed by the following strategy. pGS1.4 was digested with *Hind*III and blunt ended. The promoter insert was then

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excised with SpeI, obtaining a 1.4kb fragment with blunt-SpeI ends.

pGS1.1 was linearised with SalI and blunt ended. Later the linearised construct was digested with SpeI resulting in a linearised vector with blunt-SpeI ends containing the 3' end of the promoter region. The fragment excised in (a) was ligated into (b) to reconstruct the 2.5kb AIM-1 promoter. Figure 5 is a diagrammatic representation of the construction of the full length AIM-1 promoter.

EXAMPLE 5

Characterization of AIM-1 promoter

(a) Generation of deletion fragments and chimeric gene constructs

To fully characterise the AIM-1 promoter, two different lengths of the promoter sequence

15 were used: the entire 2.5kb sequence and a 1.4kb fragment upstream of the first ATG codon.

β-Glucuronidase (GUS) and luciferase (LUC) reporter genes were each ligated to one or

other of the promoter fragments and to the 3' terminator region from the Agrobacterium

tumefaciens nopaline synthase gene (NOS) to generate a series of chimeric gene constructs.

- A series of 7 deletions in the promoter region were also generated, starting from 170 base-pairs upstream of the first ATG codon. Each of these was likewise ligated to the NOS 3' terminator region and to the GUS reporter gene. Intermediate vectors containing each of the promoter fragments (0.17, 0.23, 0.45, 0.70, 0.88, 1.1, 1.4, 1.6 or 2.5kb) ligated to the GUS or *LUC* reporter genes and NOST were generated in pBluescript. For control purposes, additional constructs containing the cauliflower mosaic virus 35S promoter linked to either GUS or *LUC* were also prepared.
- These chimeric contructs were then successfully ligated into the polylinker of the binary vector backbone pPZP111 (Hajdukiewicz et al., 1994), for use in plant transformation. A range of these constucts, comprising the AIM-1 promoter, is shown in Figures 6A(i) to 6A(xii). The backbone vector pPZPIII is shown in Figure 6B. The pGUNt vector is shown

in Figure 6C.

(b) Transformation and regeneration of tobacco

5 The characterisation of the AIM-1 promoter was carried out using tobacco as the model plant system. Tobacco transformation was carried out as described by Svab et al. (1995).

Multiple independent transgenic lines were generated with each of the binary constructs.

(c) Generation of T2 lines

10

T2 lines were generated from selected T1 lines by self-pollination. Tissue of young transgenic tobacco lines, containing the AIM-I:GUS gene construct, were histochemically assayed to visualise GUS activity. Very intense levels of histochemical stain indicate high levels of expression of the GUS gene in tissues of young plants (Figure 7).

15

(d) Quantitative analysis of AIM-1 promoter

To quantify levels of expression of the GUS gene under control of the AIM-1 promoter and compare it to levels obtained using the CaMV35S promoter, quantitative analysis was carried out on two independent transgenic T2 tobacco lines (3-4 and 7-3) containing the AIM-1:GUS genetic construct and one transgenic T2 line (5-2) containing the 35S:GUS genetic construct. Assays were performed according to the method of Jefferson (1987) on different plant tissues including root, stem, petiole and first, second and third true leaves. The results indicated that constructs containing the AIM-1 promoter drive levels of expression 2-5 fold higher than that obtained using the 35S promoter (Figure 8).

EXAMPLE 6

Southern Analysis of Transgenic T2 Tobacco lines

Genomic DNA (10 μg) was digested with *Eco*RI or *Bam*HI restriction enzymes; separated in an electrophoresis gel and transferred to a HybondTM (Amersham) Nylon membrane. The membrane was prehybridised and hybridised at high stringency following standard procedures (Sambrook *et al.*). A DNA fragment containing the full GUS gene and Nos terminator was labelled with ³²P and used as a probe. After washing at high stringency the following results were observed:

10

- a) The *Eco*RI lanes of lines 3-4, 7-3 and 10-3 show a single fragment of the expected 4.5kb size indicating the intactness of the *AIM-1*:GUS:NosT construct in each of the above mentioned lines (see construct diagram below).
- 15 b) The BamHI lanes of lines 3-4, 7-3 and 10-3 show single fragments of different sizes (one fragment per line) indicating the existence of a single copy of GUS:NosT construct in each of the above mentioned lines.
- c) The BamHI lane of line 5-2 shows two bands indicating that the line contains two 20 copies of the CaMV 35S:GUS:NosT portion of the construct.

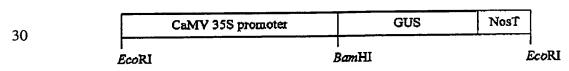
Lines 3-4, 7-3 and 10-3

2.5 kb 2.0 kb

AIM-1 promoter GUS NosT

EcoRI BamHJ EcoRI

Line 5-2



These results are shown in Figure 9.

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or more of said steps or features.

Bibliography

Botella, JR et al. Plant Mol. Biol. 20:425-436, 1992.

Botella, JR et al. Proc. Natl. Acad. Sci. USA 92:1595-1598, 1995.

Hajdukiewicz et al. Plant Mol. Biol. 25:989-994, 1994.

Jefferson, R.A. Plant. Mol. Biol. Reporter 5(4): 387-405, 1987.

Sambrook, et al. Molecular Clonging: A Laboratory Manual Cold Spring Harbor Laboratories, Cold Spring Harbor, New York, USA, 1989.

Sanger, F et al. Proc. Natl. Acad. Sci. USA 74: 5463-5467, 1977.

Selker, JML et al. Dev. Biol. 153:29-43, 1992.

Svab et al. "Methods in Plant Molecule Biology: A Laboratory Course Manual" pp55-77, Cold Spring Harbor Laboratory Press, NY, 1995.

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: THE UNIVERSITY OF QUEENSLAND
 - (ii) TITLE OF INVENTION: A NOVEL PLANT PROMOTER AND USES THEREFOR
 - (iii) NUMBER OF SEQUENCES: 9
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: DAVIES COLLISON CAVE
 - (B) STREET: 1 LITTLE COLLINS STREET
 - (C) CITY: MELBOURNE
 - (D) STATE: VICTORIA
 - (E) COUNTRY: AUSTRALIA
 - (F) ZIP: 3000
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: AUSTRALIAN PROVISIONAL
 - (B) FILING DATE: 31-AUG-1998
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: HUGHES, DR E JOHN L
 - (C) REFERENCE/DOCKET NUMBER: EJH/EK
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: +61 3 9254 2777
 - (B) TELEFAX: +61 3 9254 2770
 - (C) TELEX: AA 31787

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1923 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 88..1542

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:																	
ATCCTCTCTC CCACTTACTT CGATTTCATC AATTCCAATA AACTCAACAC ACTTTTTTAC													60				
ACTCCACACT CTAACCACAT ACACCAT ATG GGT TTC AAG GCC ATG GAC CAA Met Gly Phe Lys Ala Met Asp Gln 1														111			
ACT Thr	CCC Pro 10	TTG Leu	TTG Leu	TCC Ser	AAG Lys	ATG Met 15	GCT Ala	ATT Ile	GGG Gly	GAT Asp	GGA Gly 20	CAT His	GGC Gly	GAA Glu	TCA Ser		159
TCC Ser 25	CCA Pro	TAC Tyr	TTT Phe	GAT Asp	GGA Gly 30	TGG Trp	AAG Lys	GCT Ala	TAT Tyr	GAT Asp 35	CAA Gln	AAC Asn	CCC Pro	TTT Phe	CAT His 40		207
CCC Pro	ACA Thr	GAT Asp	AAT Asn	CCT Pro 45	AAC Asn	GGT Gly	GTT Val	ATG Met	CAA Gln 50	ATG Met	GGT Gly	CTT Leu	GCT Ala	GAG Glu 55	AAT Asn		255
CAG Gln	CTT Leu	ACC Thr	TCT Ser 60	GAT Asp	TTG Leu	GTT Val	GAA Glu	GAT Asp 65	TGG Trp	ATA Ile	CTG Leu	AAC Asn	AAC Asn 70	CCT Pro	GAA Glu		303
GCC Ala	TCC Ser	ATT Ile 75	TGC Cys	ACT Thr	CCA Pro	GAA Glu	GGA Gly 80	ATA Ile	AAT Asn	GAT Asp	TTC Phe	AGG Arg 85	GCC Ala	ATA Ile	GCT Ala		351
AAC Asn	TTT Phe 90	CAG Gln	GAT Asp	TAT Tyr	CAT His	GGT Gly 95	CTG Leu	GCC Ala	GAG Glu	TTC Phe	AGA Arg 100	AAT Asn	GCT Ala	GTG Val	GCT Ala		399
AAA Lys 105	TTT Phe	ATG Met	GCT Ala	AGA Arg	ACA Thr 110	AGG Arg	GGA Gly	AAC Asn	AGA Arg	ATC Ile 115	ACG Thr	TTT Phe	GAC Asp	CCT Pro	GAC Asp 120		447
CGT Arg	ATT Ile	GTC Val	ATG Met	AGC Ser 125	GGT Gly	GGA Gly	GCC Ala	ACC Thr	GGA Gly 130	GCA Ala	CAC His	GAA Glu	GTC Val	ACT Thr 135	GCC Ala		495
TTT Phe	TGT Cys	TTG Leu	GCA Ala 140	GAT Asp	CCC Pro	GGC Gly	GAG Glu	GCA Ala 145	Phe	TTA Leu	GTG Val	CCC Pro	ATT Ile 150	Pro	TAT Tyr		543
TAT Tyr	CCA Pro	GGC Gly 155	Phe	GAC Asp	CGG Arg	GAT Asp	TTG Leu 160	Arg	TGG Trp	AGA Arg	ACA Thr	GGA Gly 165	GTT Val	AAA Lys	CTT Leu		591
GTT Val	CCA Pro 170	Val	ATG Met	TGC Cys	GAT Asp	AGC Ser 175	Ser	AAT Asn	AAT Asn	TTC Phe	GTG Val 180	Leu	ACA Thr	AAG Lys	GAA Glu		639

GCA Ala 185	TTG Leu	GAA Glu	GAT Asp	GCC Ala	ТАТ Туг 190	GAG Glu	AAA Lys	GCA Ala	AGA Arg	GAG Glu 195	GAT Asp	AAC Asn	ATC Ile	AGA Arg	GTA Val 200	687
AAG Lys	GGT Gly	TTA Leu	CTG Leu	ATC Ile 205	ACC Thr	AAT Asn	CCA Pro	TCA Ser	AAT Asn 210	CCA Pro	TTA Leu	GGC Gly	ACA Thr	ATC Ile 215	ATG Met	735
GAC Asp	AGA Arg	AAG Lys	ACA Thr 220	CTG Leu	AGA Arg	ACC Thr	GTG Val	GTG Val 225	AGC Ser	TTC Phe	ATC Ile	AAT Asn	GAG Glu 230	AAG Lys	CGT Arg	783
ATC Ile	CAC His	CTT Leu 235	GTA Val	TGT Cys	GAT Asp	GAA Glu	ATA Ile 240	TAT Tyr	GCT Ala	GCA Ala	ACA Thr	GTT Val 245	TTC Phe	AGC Ser	CAA Gln	831
CCC Pro	GGT Gly 250	TTC Phe	ATA Ile	AGC Ser	ATA Ile	GCT Ala 255	GAG Glu	ATA Ile	TTA Leu	GAG Glu	GAT Asp 260	GAA Glu	ACA Thr	GAC Asp	ATA Ile	879
GAG Glu 265	TGT Cys	GAC Asp	CGC Arg	AAC Asn	CTC Leu 270	GTA Val	CAC His	ATT Ile	GTT Val	TAT Tyr 275	AGT Ser	CTT Leu	TCA Ser	AAG Lys	GAC Asp 280	927
ATG Met	GGG Gly	TTC Phe	CCT Pro	GGC Gly 285	TTC Phe	AGA Arg	GTC Val	GGC Gly	ATC Ile 290	ATA Ile	TAC Tyr	TCT Ser	TAC Tyr	AAT Asn 295	GAT Asp	975
GCT Ala	GTG Val	GTT Val	AAT Asn 300	TGT Cys	GCA Ala	CGC Arg	AAA Lys	ATG Met 305	TCA Ser	AGC Ser	TTT Phe	GGA Gly	TTG Leu 310	GTG Val	TCA Ser	1023
ACA Thr	CAG Gln	ACT Thr 315	CAG Gln	TAT Tyr	CTT Leu	TTA Leu	GCA Ala 320	TCG Ser	ATG Met	CTA Leu	AAT Asn	GAT Asp 325	GAT Asp	GAG Glu	TTT Phe	1071
														AGG Arg		1119
AGG Arg 345	GTT Val	TTC Phe	ACT Thr	GGG Gly	GGG Gly 350	TTG Leu	GCC Ala	AAA Lys	GTT Val	GGC Gly 355	ATA Ile	AAG Lys	TGC Cys	TTG Leu	CAA Gln 360	1167
AGC Ser	AAT Asn	GCT Ala	GGT Gly	CTA Leu 365	TTT Phe	GTG Val	TGG Trp	ATG Met	GAT Asp 370	TTA Leu	AGG Arg	CAA Gln	CTT Leu	CTC Leu 375	AAA Lys	1215
AAG Lys	CCA Pro	ACT Thr	TTC Phe 380	GAC Asp	TCT Ser	GAA Glu	ACG Thr	GAG Glu 385	CTT Leu	TGG Trp	AAA Lys	GTT Val	ATC Ile 390	ATT Ile	CAT His	1263
														ACT Thr		1311
CCA Pro	GGG Gly 410	TGG Trp	TTT Phe	AGG Arg	GTG Val	TGC Cys 415	TAT Tyr	GCC Ala	AAC Asn	ATG Met	GAT Asp 420	GAT Asp	ATG Met	GCT Ala	GTG Val	1359
														AAG Lys		1407
														AGG Arg 455		1455

AGC CTO Ser Let	AAA Lys	ACC Thr 460	AGA Arg	AGG Arg	TTT Phe	GAT Asp	GAT Asp 465	ATC Ile	ACC Thr	ATG Met	TCA Ser	CCT Pro 470	CAC His	TC:	r r	1503
CCC CTA	A CCT 1 Pro 475	Gln	TCA Ser	CCT Pro	ATG Met	GTT Val 480	AAA Lys	GCC Ala	ACA Thr	AAT Asn	TGA(GTTT(GCA			1549
TATTCC	rctg .	AATC	TTTF	AG AZ	GAAC	TAAC	TG	TATO	TGA	AGAT	TAC	rtg (GTTC	rtt:	TAT	1609
TTGTTA!	TTTT	GAGA	AGGT	AC AT	raagi	rgcTc	GAT	TTGT	TCT	TTGG	SAAC	AGC 2	ATAA	ACA	GGA	1669
AATTCC	rgat	GTTGI	TTTT	et G	ATCGO	CATO	ACA	ATCO	CAGT	GTCC	TAC	AAG '	rtgt	GCT(GCT	1729
TCATGC	ACGC	CCCTI	CAAT	rc Ti	raggo	GCAT	r TTT	TTCI	TTT	TTCA	CTT	ACC I	AAAG	GTT(CAA	1789
GGTGAA	AAAA	GTTTA	TAG	G TO	CTGT	\ATGT	TAT	TGGT	ATTT	TCAG	AAG	AGT (CCAA	AAG	ATG	1849
TCTGTA	ATCT	GCTAC	TGA	A T	GTA	CTTI	CAZ	ATTA	rgaa	TAAA	TTG:	TA Z	AATA	AGG'	TCT	1909
TCAAAT'	CAT	TTCC														1923

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 484 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Gly Phe Lys Ala Met Asp Gln Thr Pro Leu Leu Ser Lys Met Ala Ile Gly Asp Gly His Gly Glu Ser Ser Pro Tyr Phe Asp Gly Trp Lys 20 25 30 Ala Tyr Asp Gln Asn Pro Phe His Pro Thr Asp Asn Pro Asn Gly Val Met Gln Met Gly Leu Ala Glu Asn Gln Leu Thr Ser Asp Leu Val Glu Asp Trp Ile Leu Asn Asn Pro Glu Ala Ser Ile Cys Thr Pro Glu Gly 65 70 75 Ile Asn Asp Phe Arg Ala Ile Ala Asn Phe Gln Asp Tyr His Gly Leu Ala Glu Phe Arg Asn Ala Val Ala Lys Phe Met Ala Arg Thr Arg Gly 100 105 110 Asn Arg Ile Thr Phe Asp Pro Asp Arg Ile Val Met Ser Gly Gly Ala Thr Gly Ala His Glu Val Thr Ala Phe Cys Leu Ala Asp Pro Gly Glu 135 Ala Phe Leu Val Pro Ile Pro Tyr Tyr Pro Gly Phe Asp Arg Asp Leu 145 150 155 160

Arg Trp Arg Thr Gly Val Lys Leu Val Pro Val Met Cys Asp Ser Ser

Lys Ala Thr Asn

Asn Asn Phe Val Leu Thr Lys Glu Ala Leu Glu Asp Ala Tyr Glu Lys Ala Arg Glu Asp Asn Ile Arg Val Lys Gly Leu Leu Ile Thr Asn Pro Ser Asn Pro Leu Gly Thr Ile Met Asp Arg Lys Thr Leu Arg Thr Val 215 Val Ser Phe Ile Asn Glu Lys Arg Ile His Leu Val Cys Asp Glu Ile Tyr Ala Ala Thr Val Phe Ser Gln Pro Gly Phe Ile Ser Ile Ala Glu Ile Leu Glu Asp Glu Thr Asp Ile Glu Cys Asp Arg Asn Leu Val His 265 Ile Val Tyr Ser Leu Ser Lys Asp Met Gly Phe Pro Gly Phe Arg Val Gly Ile Ile Tyr Ser Tyr Asn Asp Ala Val Val Asn Cys Ala Arg Lys Met Ser Ser Phe Gly Leu Val Ser Thr Gln Thr Gln Tyr Leu Leu Ala 305 Ser Met Leu Asn Asp Asp Glu Phe Val Glu Arg Phe Leu Ala Glu Ser Ala Lys Arg Leu Ala Gln Arg Phe Arg Val Phe Thr Gly Gly Leu Ala Lys Val Gly Ile Lys Cys Leu Gln Ser Asn Ala Gly Leu Phe Val Trp Met Asp Leu Arg Gln Leu Leu Lys Lys Pro Thr Phe Asp Ser Glu Thr Glu Leu Trp Lys Val Ile Ile His Glu Val Lys Ile Asn Val Ser Pro Gly Tyr Ser Phe His Cys Thr Glu Pro Gly Trp Phe Arg Val Cys Tyr 410 Ala Asn Met Asp Asp Met Ala Val Gln Ile Ala Leu Gln Arg Ile Arg 425 Asn Phe Val Leu Gln Asn Lys Glu Val Val Ser Asn Lys Lys His Cys Trp His Ser Asn Leu Arg Leu Ser Leu Lys Thr Arg Arg Phe Asp 455 Asp Ile Thr Met Ser Pro His Ser Pro Leu Pro Gln Ser Pro Met Val 470 475

2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2474 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

60	AATAAGTGTC	ACAGAAAAAT	TACTTTAATA	ACGACACATC	ACAGAATCAG	TTACAGATAC
120	TTGGTCTGTT	TGAGATTATT	TTTATATGGT	ATGAAATGTT	GTACGACAAG	GGAGATTATG
180	AAGTTGAATC	AAAAAATGAA	TTTACGTATT	TGATTTTGAT	TCACGAATCA	GTTGGAAGTT
240	ACAACTGCAG	AGCCAGTTGA	CCAAAAAAAT	GGGAACTGAA	TCTAGAAGCT	ATGCATTTTA
300	CTTTATCGGA	CTTGGTTGCT	TACAATAATC	TCTCCTTTCC	CGTATTCATT	TATTTGTAGG
360	AACATCACAT	TGCCGACAAG	GGTCCTCGAT	TACTCTGTAA	AAGCAATAGC	AAAAAACCAA
420	TTTGCATATT	ACGTGCGAGT	GTTGAAGCTC	TAATTTTGAG	GAAGAACACA	GCGTGCTGTC
480	TAGTTGAGTG	ATAGTGTAGG	TTCCGCGTAT	ATGGAGTGAG	GTGTACACGT	TTTAGGTTAT
540	AAGGGATTTA	CCACCTAATT	TCTTTTCAAG	GGTAACACTA	GAGTGAATCA	GCTGAGTAGC
600	TATTCTCTCT	ACCTTTATAA	GGCCCCACTT	CGCTAACTAA	AACTGTTCTT	ATGTTCATGC
660	GATTATTTT	TTAATGGAGG	CTACTCTTAT	TACAACTTTT	TTTTGGTAAG	AACTCCGGGC
720	TAAAACAATT	GATCTTATAT	TATGCATTTT	TTTTTTAATT	AATTAATTTA	TCCATATACC
780	TTAGCCATTT	TTCCTCAAGT	AATTGAAGTT	TATCGGTGAC	TTAAGTCGTA	ATGGTATGGA
840	TAACAATTTC	TATGTATCAT	GGTAAATTCA	ACTACTATTA	AAACTTAATC	TTATGAAATT
900	TAACTAAAGT	CAACTTCTGT	AAGTTGTTGT	CAAGATTTGA	CAATTTTACC	AATGTGAGTT
960	GTTGATGACT	AATTGAAGGG	ATCTATTTTG	TTTAACCTAA	GGTTGACGAC	TGTATTATAA
1020	GTGTTCATAA	GGAGATTTTA	AGACTACATT	CTAAAGTTCT	AATAATTCAA	TCAGCTTTAA
1080	CAATTAATTA	GGTGACTATT	GAAAAAGATT	TTAAAGTTAT	AAAGGCTGAG	AATTTTAGAA
1140	CCACCTCGAC	CAGAGAAATA	CATAACCAAT	ATTTCATGAG	GATGACAAAT	GTTGTGAATT
1200	TCTAGAATAA	GTATAAGGAG	AAGCATTGTA	TTAATTAATG	AATCTCAATG	CGACTACAAC
1260	TTGATCTATA	CAAGAAAAGT	ТАААААТТА	AAACTATTTT	TATTAGAGGA	ATTTCTTAAA
1320	GTTGATAGGG	CTCACATTGT	TTTCTTATGA	ATCTAACAAT	ACTTTAAATT	ACCTCTTTAA
1380	AATTATATAT	TTTGTCTGCG	ACTAGTATGA	TCTATTTTAT	AAAATATATG	TGATTTTGTC
1440	ATATTTATTC	AAAGGAGAAA	AAGTTATAAA	ATTGCCTAAT	TGGAGAAATG	AGTATTAACT
1500	GAGATTAAGA	САТТАТАТАА	ТААТААААА	TAAGTAACAA	TACACTTAAA	АТААААААА
1560	TAAAATATTC	AAATTTGAAC	TTTTATTAT	GTAGAATAAT	AGTATTGAAT	ТААТТТААТА
1620	AATTCAATCT	CAATACGAGT	ATTCATCATT	AATAGATATA	CAAAGTAAAT	TTATAATAAA

ATTATAATCA	TATATTAGAT	AAATATACAA	ATATTTGTTA	AATTTTACAT	TATTATATTA	1680
СТАААТАТАТ	ATTAATTTCT	TTGAATATCT	TTTATACAAG	TAGGTAGACT	AGAAGAATTA	1740
TCTTATCTCC	CGTATATTTG	TAGATGTTAA	ATGTAACGGG	CTTAGACTGA	TGTTTTTGTA	1800
TTATATTATT	TATAAATCCA	TTAGAGATTT	AAGTTAATGT	CTCTCTTTGA	TTTTAACATG	1860
GTTCTAAAAA	TTAGGTTTAA	TCATTGCGTC	CTCAATGAAC	CCATGCTATA	TGTTTTAAAG	1920
TTTTTTGTTT	TTTGACAATG	TTTTTTTTTT	CTGAGATTGC	TCTTAGGATT	GAAATTATGT	1980
TTGATACTAG	AAAACGAAGA	AGTAGAGAGT	AGTGTATACA	CGTGTAAAAA	ATAATAGTTG	2040
TGGGAACTTA	AGTTGGATTT	GAATACTAGG	ACGAGGCTGG	AAGGGTTTCC	ACTAAGTTGA	2100
CAAAAATTAT	TACAAGTGGC	AACTAGCTAG	GTCTCACAAA	GTATTACTAA	TTAATAGTGG	2160
GTCTGTCTGC	ATACCAACTC	TTGCCTAATT	TTCAAACACC	GCATTCTCTC	TTCTTCTCTC	2220
CTTCTTCCTC	TGGAAACTTC	ATCGATGTGG	ACTTCTGTCT	CTCAAAAGTC	AAGCTCAATT	2280
TATCCAATGC	ATTATAAATA	CACACTCTCC	CTCCCTTCTA	TTCTTCATTG	CATCACATTT	2340
CCTCTATAAA	TTACTCACAC	CTTATTCCTA	ACTTCATTTC	AACATCCTCT	CTCCCACTTA	2400
CTTCGATTTC	ATCAATTCCA	АТАААСТСАА	CACACTTTTT	TACACTCCAC	ACTCTAACCA	2460
CATACACCAT	ATGG					2474

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4: GCGGATCCAT CTTGGACAAC AAGGGAGTT

2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 30 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5: TAGGATCCAG AAAGACACTG AGAACCGTGG	30
(2) INFORMATION FOR SEQ ID NO:6:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA	
(xi) ·SEQUENCE DESCRIPTION: SEQ ID NO:6:	
ACGGATCCGG TGTATGTGGT TAGAGTGTG	29
(2) INFORMATION FOR SEQ ID NO:7:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
CAGGATCCAG ACATAGAGTG TGACCGCAA	29
(2) INFORMATION FOR SEQ ID NO:8	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 59 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8	

ATCGATCATA TGAGCTCTAG ACCCGGGCTG CAGGATCCGG TGTATGTGGT TAGAGTGTG

- (2) INFORMATION FOR SEQ ID NO:9
 - (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 57 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9

CCGCGGAGAT CTATCGATCT CGAGAATTCA AGCTTCAGAC ATAGAGTGTG ACCGCAA

57

DATED this 31st day of August, 1998

THE UNIVERSITY OF QUEENSLAND by Its Patent Attorneys DAVIES COLLISON CAVE

AIM-1 OLIGONUCLEOTIDES

-Oligonucleotide primers used during Long Distance Inverse PCR -Oligo's bind to regions of AIM-1 (Mungbean ACC Synthase).

NSE-1

5` -GCGGAT[↓]CCATCTTGGACAACAAGGGAGTT- 3` 29'omer

Tm = 68

NSE-2

5' -TAGGATC[↓]CAGAAAGACACTGAGAACCGTGG- 3'

30'omer

Tm = 70

NSE-3

5' -ACGGATCC[↓]GGTGTATGTGGTTAGAGTGTG- 3'

29'omer

Tm = 62

NSE-4

5`-CAGGATC[↓]CAGACATAGAGTGTGACCGCAA- 3`

29'omer

Tm = 66

NSE-5

5'-ATCGATCATATGAGCTCTAGACCCGGGCTGCAGGATCC GGTGTATGTGGTTAGAGTGTG- 3'
59'omer

Tm = 62

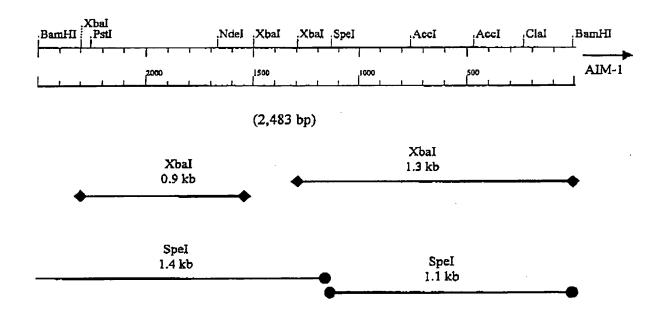
note: NSE-5 is identical to NSE-3 except different restriction enzyme sites have been incorporated (ie. 5'-Cla I, Nde I, Sac I, Xba I, Sma I, Pst I & Bam H1-3')

NSE-6

5'-CCGCGGAGATCTATCGATCTCGAGAATTC<u>AAGCTT</u> ↓CAGACATAGAGTGTGACCGCAA-3' 57'omer

Tm = 66

note: NSE-6 is identical to NSE-4 except different restriction enzyme sites have been incorporated (ie. 5'-Sac II, Bgl II, Cla I, Xho I, Eco R1, & Hind III-3')



AIM-1 2.5Kb promoter sequencing strategy:

```
All fragments
Xba-14F
                                 <----+
Spe-12R
                             <----+
Spe-10F
Spe-09F
Spe-06R
Spe-10R
Spe-12F
Spe-06F
Spe-03
Xba-14R
Spe-14F
            <----+
Xba-09F3
         <----+
Xba-09F
       +---->
Xba-09R
     +---->
CONSENSUS+---->
Spe-1.4
     0 300 600 900 1200 1500 1800 2100 2400
```

FIGURE 3

IM-1 2.5Kb Promoter French

Length: 2470

1	TTACAGATAC	ACAGAATCAG	ACGACACATC	TACTTTAATA	ACAGAAAAAT
51	AATAAGTGTC	GGAGATTATG	GTACGACAAG	ATGAAATGTT	TTTATATGGT
101	TGAGATTATT	TTGGTCTGTT	GTTGGAAGTT	TCACGAATCA	TGATTTTGAT
151	TTTACGTATT	AAAAAATGAA	AAGTTGAATC	ATGCATTTTA	TCTAGAAGCT
201	GGGAACTGAA	ССААААААТ	AGCCAGTTGA	ACAACTGCAG	TATTTGTAGG
251	CGTATTCATT	TCTCCTTTCC	TACAATAATC	CTTGGTTGCT	CTTTATCGGA
301	AAAAAACCAA	AAGCAATAGC	TACTCTGTAA	GGTCCTCGAT	TGCCGACAAG
351	AACATCACAT	GCGTGCTGTC	GAAGAACACA	TAATTTTGAG	GTTGAAGCTC
401	ACGTGCGAGT	TTTGCATATT	TTTAGGTTAT	GTGTACACGT	ATGGAGTGAG
451	TTCCGCGTAT	ATAGTGTAGG	TAGTTGAGTG	GCTGAGTAGC	GAGTGAATCA
501	GGTAACACTA	TCTTTTCAAG	CCACCTAATT	AAGGGATTTA	ATGTTCATGC
551	AACTGTTCTT	CGCTAACTAA	GGCCCCACTT	ACCTTTATAA	TATTCTCTCT
601	AACTCCGGGC	TTTTGGTAAG	TACAACTTTT	CTACTCTTAT	TTAATGGAGG
651	GATTATTTTT	TCCATATACC	AATTAATTTA	TTTTTTAATT	TATGCATTTT
701	GATCTTATAT	TAAAACAATT	ATGGTATGGA	TTAAGTCGTA	TATCGGTGAC
751	AATTGAAGTT	TTCCTCAAGT	TTAGCCATTT	TTATGAAATT	AAACTTAATC
801	ACTACTATTA	GGTAAATTCA	TATGTATCAT	TAACAATTTC	AATGTGAGTT
851	CAATTTTACC	CAAGATTTGA	AAGTTGTTGT	CAACTTCTGT	TAACTAAAGT
901	TGTATTATAA	GGTTGACGAC	TTTAACCTAA	ATCTATTTTG	AATTGAAGGG
951	GTTGATGACT	TCAGCTTTAA	AATAATTCAA	CTAAAGTTCT	AGACTACATT
1001	GGAGATTTTA	GTGTTCATAA	AATTTTAGAA	AAAGGCTGAG	TTAAAGTTAT
1051	GAAAAAGATT	GGTGACTATT	CAATTAATTA	GTTGTGAATT	GATGACAAAT
1101	ATTTCATGAG	CATAACCAAT	CAGAGAAATA	CCACCTCGAC	CGACTACAAC
1151	AATCTCAATG	TTAATTAATG	AAGCATTGTA	GTATAAGGAG	TC'IAGAATAA
1201	ATTTCTTAAA	TATTAGAGGA	AAACTATTTT	TAAAAAATTA	CAAGAAAAGT
1251	TTGATCTATA	ACCTCTTTAA	ACTTTAAATT	ATCTAACAAT	TTTCTTATGA

1301	CTCACATTGT	GTTGATAGGG	TGATTTTGTC	AAAA ATG	TCTATTTTAT
1351	ACTAGTATGA	TTTGTCTGCG	AATTATATAT	AGTATTAACT	TGGAGAAATG
1401	ATTGCCTAAT	AAGTTATAAA	AAAGGAGAAA	ATATTTATTC	ATAAAAAAA
1451	TACACTTAAA	TAAGTAACAA	ТААТАААААА	CATTATATAA	GAGATTAAGA
1501	TAATTTAATA	AGTATTGAAT	GTAGAATAAT	TTTTATTTAT	AAATTTGAAC
1551	TAAAATATTC	AAATAATATT	CAAAGTAAAT	AATAGATATA	ATTCATCATT
1601	CAATACGAGT	AATTCAATCT	ATTATAATCC	ATATATTAGA	TAAATATACA
1651	AATATTTGTT	AAATTTTACA	TTATTATATT	ACTAAATATA	TATTAATTTC
1701	CTTTGAATAT	CTTTTATACA	AGTAGGTAGA	CTAGAAGAAT	TATCTTATCT
1751	CCCGTATATT	TGTAGATGTT	AAATGTAACG	GGCTTAGACT	GATGTTTTTG
1801	TATTATATTA	TTTATAAATC	CATTAGAGAT	TTAAGTTAAT	GTCTCTCTTT
1851	GATTTTAAAC	ATGGTCTAAA	AATTAGGTTT	AATCATTGCG	TCCTCAATGA
1901	ACCCATGCTA	TATGTTTTAA	AGTTTTTTGT	TTTTTGACAA	TGTTTTTTAT
1951	TTCTGAGATT	GCTCTTAGGA	TTGAAATTAT	GTTTGATACT	AGAAAACGAA
2001	GAAGTAGAGA	GTAGTGTATA	CACGTGTAAA	AAATAATAGT	TGTGGGAACT
2051	TAAGTTGGAT	TTGAATACTA	GGACGAGGCT	GGAAGGGTTT	CCACTAAGTT
2101	GACAAAAATT	ATTACAAGTG	GCAACTAGCT	AGGTCTCACA	AAGTATTACT
2151	AATTAATAGT	GGGTCTGTCT	GCATACCAAC	TCTTGCCTAA	TTTTCAAACA
2201	CCGCATTCTC	TCTTCTTCTC	TCCTTCTTCC	TCTGGAAACT	TCATCGATGT
2251	GGACTTCTGT	CTCTCAAAAG	TCAAGCTCAA	TTTATCCAAT	GCATTATAAA
2301	TACACACTCT	CCCTCCCTTC	TATTCTTCAT	TGCATCACAT	TTCCTCTATA
2351	AATTACTCAC	ACCTTATTCC	TAACTTCATT	TCAACATCCT	CTCTCCCACT
2401	TACTTCGATT	TCATCAATTC	CAATAAACTC	AACACACTTT	TTTACACTCC
2451	ACACTCTAAC	CACATACACC			

FIGURE 4 c ntinued

Recor iction of 2.5 kb AIM-1 probe

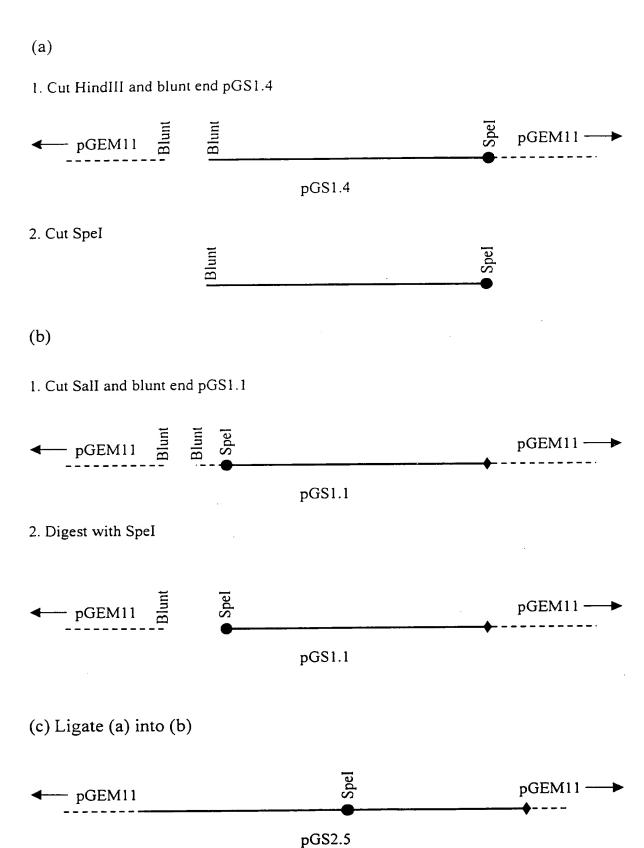
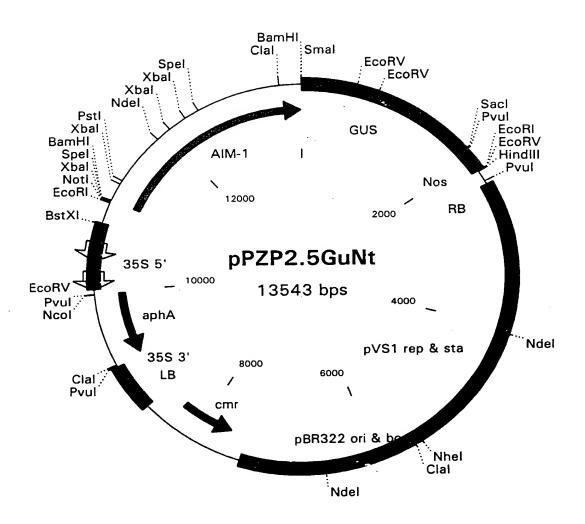
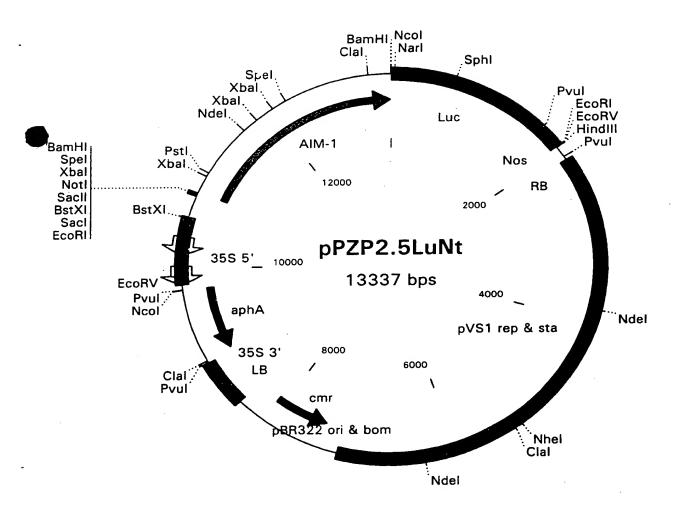


FIGURE 5





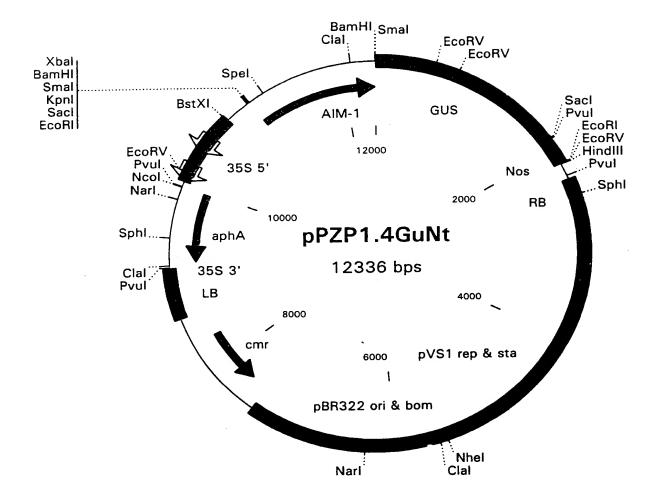


FIGURE 6A(iii)

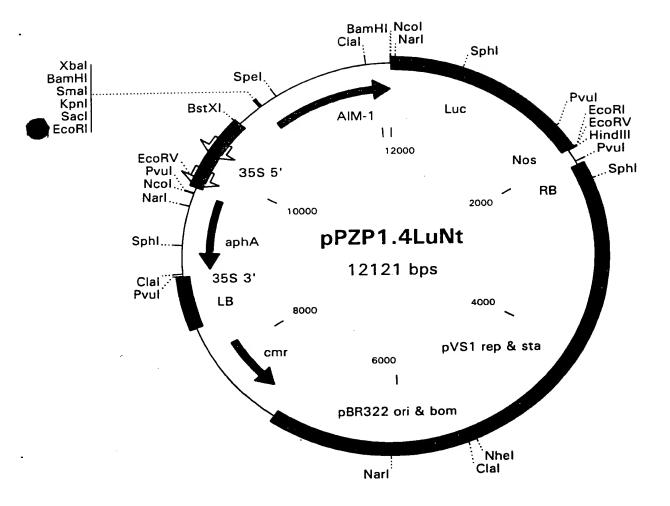


FIGURE 6A(iv)

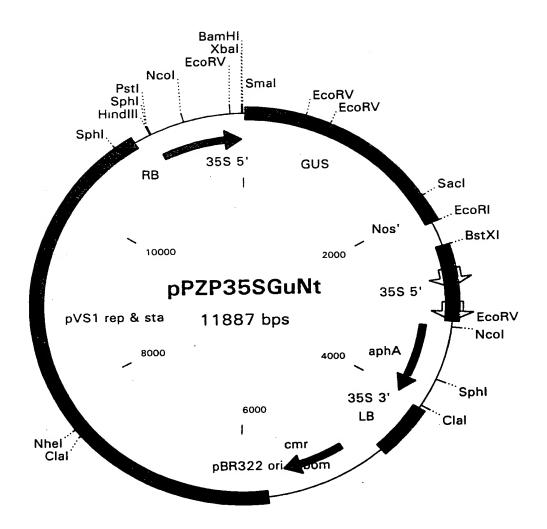


FIGURE 6A(v)

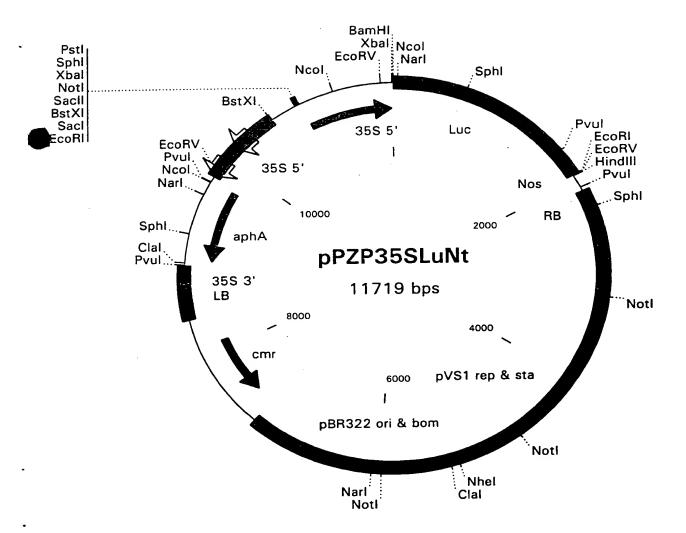


FIGURE 6A(vi)

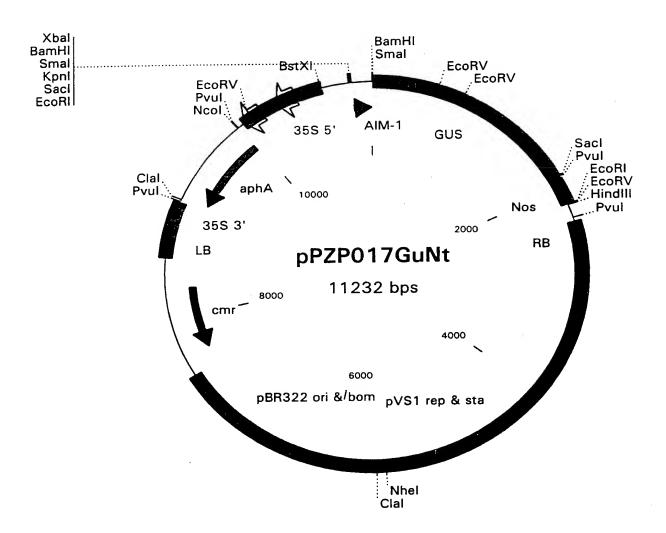
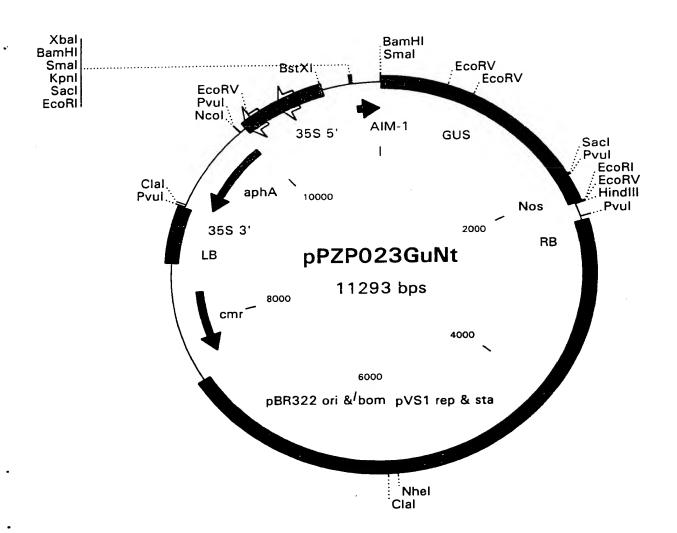


FIGURE 6A(vii)



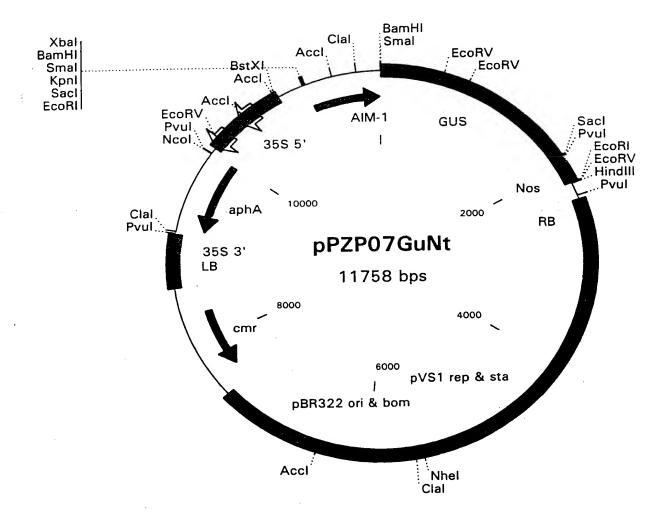
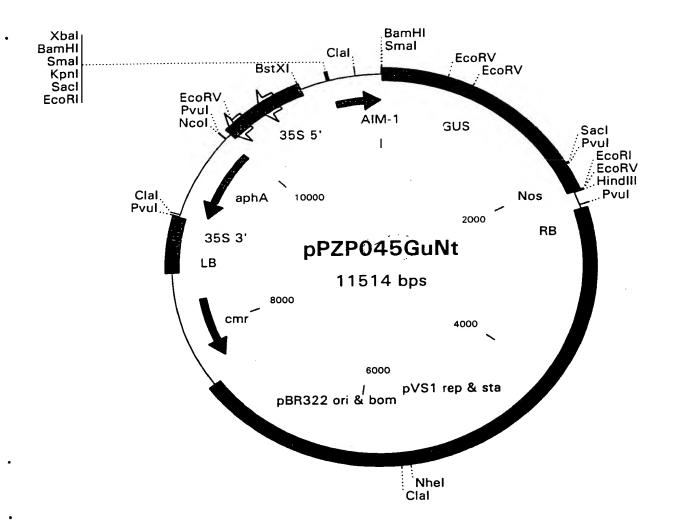


FIGURE 6(ix)



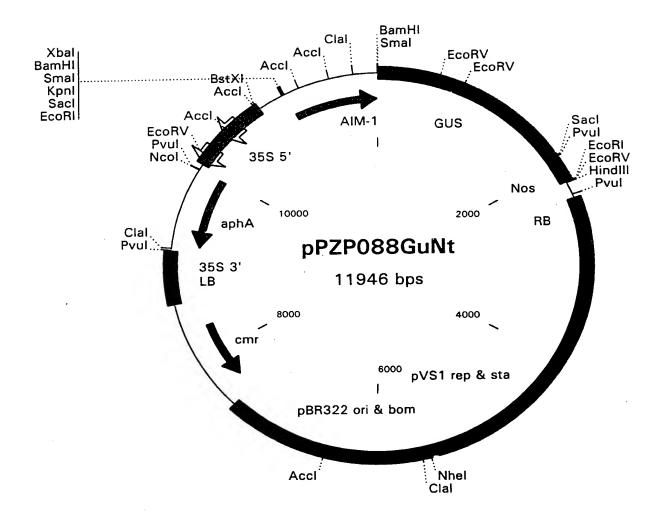


FIGURE 6A(xi)

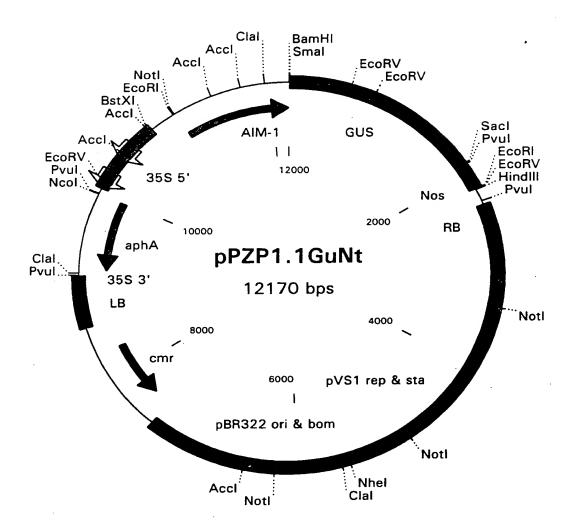


FIGURE 6A(xii)

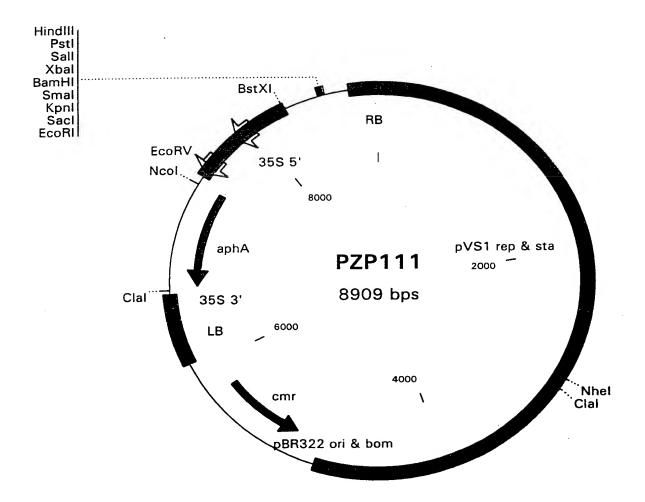


FIGURE 6B

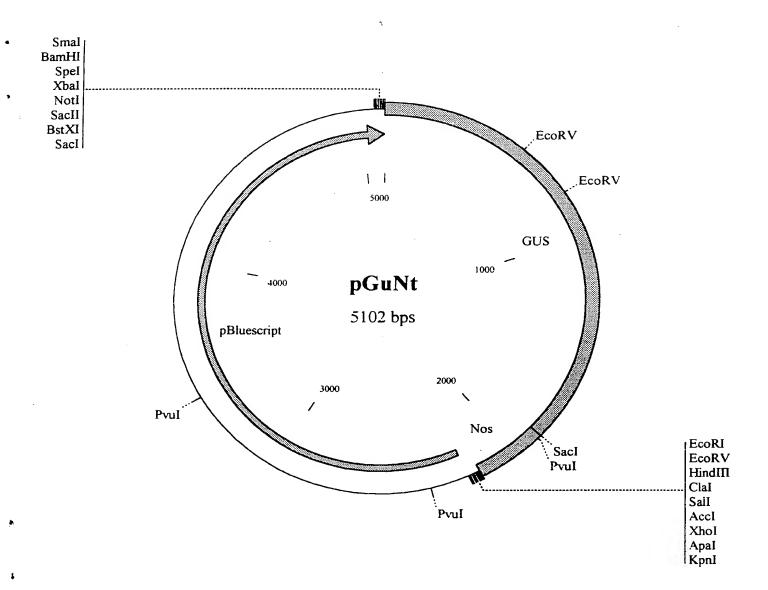


FIGURE 6C

FIGURE 7